This article was downloaded by: On: 24 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



**To cite this Article** Cserháti, T. , Szögyi, M. and Lelkes, L.(1995) 'Charge-Transfer Chromatography Used To Study The Interaction of Chlorhexidine with Proteins and Amino Acids', Journal of Liquid Chromatography & Related Technologies, 18: 11, 2175 – 2192

To link to this Article: DOI: 10.1080/10826079508010263 URL: http://dx.doi.org/10.1080/10826079508010263

Taylor & Fra

# PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

# CHARGE-TRANSFER CHROMATOGRAPHY USED TO STUDY THE INTERACTION OF CHLORHEXIDINE WITH PROTEINS AND AMINO ACIDS

T. CSERHÁTI<sup>1</sup>, M. SZÖGYI<sup>2</sup>, AND L. LELKES<sup>3</sup>

<sup>1</sup>Central Research Institute for Chemistry Hungarian Academy of Sciences P.O. Box 17, 1525 Budapest, Hungary <sup>2</sup>Institute of Biophysics Semmelweis Medical University Budapest, Hungary <sup>3</sup>Central Pharmacy Visegrad, Hungary

#### ABSTRACT

The interaction of the antibacterial agent chlorhexidine (1,1-hexamethylene-bis-/5-(4-chlorophenyl)-bisguanidine/) with human albumin and pepsin and with amino acids was studied with reversed-phase charge transfer chromatography in the presence of various monovalent cations. It was established that the mobility of chlorhexidine increases in the presence of ions in the lower concentration range (0.001 - 0.1 M) then increases at higher concentrations. The ion radii significantly influences the effect. Stepwise regression analysis proved that the chlorhexidine binds both to pepsin and human albumin, the binding is stronger with human albumin. The ions modify the character of chlorhexidine-human albumin binding indicating the hydrophilic character of the interaction. Except Asp and

# 2175

Copyright @ 1995 by Marcel Dekker, Inc.

Glu the other amino acids exhibited weak interactions with chlorhexidine, no significant difference was found between the interactive strength of L- and Dforms of Asp and Glu. Our data make probable that hydrophilic binding of chlorhexidine to the acidic side chains in proteins may be involved in its mode of action.

#### INTRODUCTION

Many studies have been devoted to the exploration of the effect of chlorhexidine (1,1-hexamethylenebis-/5-(4-chlorophenyl)-bis-guanidine/) on the reduction of gingivitis (1-6) and plaque (7-12) formation. Chlorhexidine effectively reduces oral microflora (13-23) and the incidence of oral mucosal complications after bone marrow transplantation (24), inactivates human immunodeficiency virus (25), inhibits the incorporation in DNA of thymidine, decreases lactate dehydrogenase content in human buccal epithelial cells in culture (26) and desorbs adsorbed salivary constituents (27). However, chlorhexidine causes burns of the lips, mouth and tongue (28) and anaphylactic symptoms (29). Chlorhexidine interacts with surfactants (30), the addition of Triton-X-100 (31) or sodium lauryl sulphate (32) decreased its effect.

The mode of action of chlorhexidine has not been elucidated in detail. Chlorhexidine is thought to interact with acidic lipid components to cause changes in the membrane permeability (33), however, it changes only slightly the phase transition temperature of phosphatidylcholine and phosphatidylglycerol (34).

Charge-transfer reversed-phase thin-layer chromatography has been frequently used to study interactions between different organic molecules of low molecular weight. The theory and practice of the determination of relative interactive strength by this method has also been described (35).

The objectives of our work were to study the interaction of chlorhexidine with some proteins and amino acids and to elucidate the effect of ions on the strength of interaction.

## MATERIALS AND METHODS

Chlorhexidine diacetate (Serva Chemical Co.. USA) was of analytical purity. Human albumin and pepsin were the products of Human Vaccine Works (Budapest, Hungary) and Chemical Works of Gedeon Richter Ltd. (Budapest, Hungary), respectively. Amino acids were of analytical purity and of L-conformation, the D-forms were included in the experiments only in the case of Asp and Glu. To study the interaction of chlorhexidine with the proteins cellulose powder for TLC (Merck, Darmstadt, Germany) was mixed with the proteins in 9:1, 19:1 and 99:1 w/w ratios. Layers of 20 x 20 cm (0.25 mm thickness) were prepared from the mixed slurries and after drying their UV spectra was determined with a Model CS-930 Dual Wavelength TLC Scanner (Shimadzu, Kyoto, Japan). The UV spectra of chlorhexidine adsorbed on cellulose layer was determined in separate experiment. As the chlorhexidine was easily detectable even at the highest protein:cellulose ratio (1:9 w/w), this mixed sorbent was used in the experiments to study the interaction between chlorhexidine and the proteins. The use of unimpregnated cellulose layer as reversed-phase sorbent was motivated by the theoretical considerations stating that any layer may behave as a reversed-phase one when the stationary phase is less polar than the mobile phase (36). The validity of the hypothesis outlined above was proved to be true for unimpregnated cellulose (37). Chlorhexidine diacetate was dissolved in distilled water at the concentration of 2 mg/ml, 5  $\mu$ l of this solution was spotted onto the plates. Distilled water was used as eluent containing LiCl, NaCl and KCl in the concentration range of 0.001 - 1 mM. At each salt concentration unmixed cellulose layers served as control. After development the plates were dried at 105°C and the maximum of the chlorhexidine spot was determined with the same TLC scanner at 270 nm. The R<sub>M</sub> values characterizing the lipophilicity were calculated in each case.

As it was assumed that the protein and salt concentration and the interaction between proteins and salts may influence simultaneously the  $R_M$  value of chlorhexidine we used stepwise resression analysis to elucidate this problem (38). As the exact type of correlation (linear, quadratic or logarithmic) between the independent and dependent variables was not previously established we used the following independent variables:  $x_1$  = human albumin content of the sorbent (%);  $x_2$  = pepsin content of the sorbent (%);  $x_3$  = cation radii;  $x_4$  = (log  $x_6$ )<sup>2</sup>;  $x_5$  = ( $x_1$ . $x_7$ )<sup>2</sup>;  $x_6$  = ( $x_7$ )<sup>2</sup>;  $x_7$  = salt concentration in the eluent (M);  $x_8$  =  $x_1$ . $x_7$ ;  $x_9$  =  $x_2$ . $x_7$ ;  $x_{10}$  = ( $x_2$ . $x_7$ )<sup>2</sup>;  $x_{11}$  = log  $x_7$ . The number of accepted variables was not limited, the significance level for each independent variables was set to 95%.

To study the interaction of chlorhexidine with amino acids DC-Alufolien cellulose plates (Merck, Darmstadt, Germany) were used without any pretreatment. Chlorhexidine was spotted onto the plates as described above. For the screening test distilled water was used as the eluent containing the amino acids at 10 mM concentration. The detection of the spot maximum was carried out by the TLC scanner as described above. As Asp and Glu showed the highest effect their interaction with chlorhexidine was studied more in detail, the concentrations of Asp and Glu varying between 0 -10 and 0 - 100 mM depending on their solubility in water. To elucidate the role of stereospecificity the experiments with Asp and Glu were carried out with the L- and D-forms too. As the correlation between the Asp and Glu concentration and the  $R_M$  value of chlorhexidine seemed to be markedly nonlinear, logarithmic correlations were calculated between the  $R_M$  value and amino acid concentration separately for the L- and Dforms of Asp and Glu.

### RESULTS AND DISCUSSION

The UV spectra of chlorhexidine and human albumin adsorbed on cellulose are shown in Fig.1. The chlorhexidine has an UV maximum at 270 nm. The human albumin (and also pepsin) has a relatively low absorbance at this wavelength, therefore the adsorbed proteins interfere to a negligible extent with the UV detection of chlorhexidine.

The lipophilicity of chlorhexidine decreasad rapidly with the growing concentration of salt in the eluent (Fig.2) then - having a minimum - increased again at higher salt concentrations. However, this increasing phase was slower than the decreasing phase. The effect was similar for each salt but depended also



FIGURE 1. UV spectra of chlorhexidine (A) and human albumin (B) adsorbed on cellulose. I. human albumin: cellulose 1.9 w/w; II. human albumin:cellulose 1:19 w/w; III. human albumin:cellulose 1:99 w/w



FIGURE 2. Effect of various salts on the  $\rm R_{M}$  value of chlorhexidine.

on the type of salt. Our data make obvious that the salt in the eluent influences the  $R_M$  value of chlorhexidine in minimally two different ways. We assume that the polar groups of chlorhexidine interact with the polar adsorptive site of cellulose. The ions present in the eluent also bind to the polar adsorption centers of cellulose. This coadsorption is of competitive character, the higher the ion strength the higher is the ratio of adsorptive site occupied by the ions in the eluent. The lessening number of binding sites results in continuous decrease of chlorhexidine retention. Similar masking effects of eluent additives was also observed in the reversed-phase chromatography of peptides (39). After the probable saturation of free adsorptive sites of cellulose by the ions in the eluent the situation changes markedly. The fact that higher salt concentrations increase again the  $R_M$  values of chlorhexidine may be due to the dissociation suppressing effect of salts. The undissociated chlorhexidine molecule shows a higher reversed-phase retention than the dissociated one. Similar anomalous effect of salts on the reversed-phase retention behavior of heterocyclic quaternary ammonium salts has been previously reported (40).

The proteins markedly decreased the retention of chlorhexidine at higher salt concentration (Fig.3) indicating that the proteins really interact with chlorhexidine.

The results of stepwise regression analysis are compiled in Table 1.

The equation fits well to the experimental data the significance level being over 99.9% (compare calcula-ted and tabulated F values). Six independent variables



FIGURE 3. Effect of human albumin (1) and pepsin (2) on the lipophilicity of chlorhexidine.

of the eleven included ones account for about 74% of the total variance (see  $r^2$  value). It can be concluded from the slope (b) values that both human albumin and pepsin significantly increase the retention of chlorhexidine that proves the binding of chlorhexidine to these proteins. The ions with greater ion radii decrease more strongly the retention of chlorhexidine that is not only the ion concentration but also the character of the cation influences the lipophilicity

TABLE 1. Effect of Salts and Proteins on the Lipophilicity  $(R_{\mu})$ of Chlorhexidine. Results of Stepwise Regression Analysis.  $R_M = a + b_1 \cdot x_1 + b_2 \cdot x_2 + b_3 \cdot x_3 + b_4 \cdot (\log x_7)^2 + b_5 \cdot x_1 \cdot x_7$  $+ b_{6} \cdot (x_{7})$ n = 74 a = 0.52  $F_{calc} = 32.59$   $F_{99.9\%} = 4.37$   $r^2 = 0.7420$ Number of indepen-  $b.10^2$ s<sub>b</sub>.10<sup>3</sup> Path coefficient % dent variable 1 2.10 5.16 12.4 2 1.27 5.26 6.0 3 -21.97 10.73 5.0 4 10.55 8.50 33.5 5 -0.46 1.28 11.9 6 74.72 75.07 30.9  $x_1$  = human serum albumin content of the sorbent (%)  $x_2$  = pepsin content of the sorbent (%)  $x_3 = cation radii$ 

 $x_7$  = salt concentration in the eluent (M)

of chlorhexidine. The exact type of correlation between the salt concentration in the eluent and the  $R_M$ value of chlorhexidine is not clear. Our data suggest that it can be quadratic or log quadratic. This finding supports our previous assumption that the influence of salts is composed of more than one physicochemical processes which are yet not clearly understood. The interaction between human albumin and salt concentration decreases the  $R_M$  value of chlorhexidine proving again the hydrophilic character of proteinchlorhexidine interaction. The path coefficients show the relative impact of the independent variables on the  $R_M$  value. The salt concentration exerts the highest effect on the retention followed by the proTABLE 2. Effect of Amino Acids on the Lipophilicity of Chlorhexidine.

Lipophilicity (R <sub>M</sub> n Standard deviation	value) Coefficient of variation%
<b>0.03</b>	2.57
0.03	2.41
0.04	3.88
0.02	1.91
8 0.03	11.81
0.02	1.41
0.03	5.47
5 0.02	1.64
5 0.08	7.35
0.01	0.52
0.10	9.30
0.05	4.18
0.02	2.24
5 0.02	1.54
0.03	2.57
0.05	3.57
0.02	1.21
0.04	3.10
	Lipophilicity (R <sub>M</sub> Standard deviation 0.03 0.03 0.04 0.02 0.02 0.02 0.02 0.03 0.03 0.03 0.03

tein concentrations. The impact of the ion radii is of secondary importance.

The results of the screening test for amino acidchlorhexidine interaction are compiled in Table 2. Many amino acids decrease the lipophilicity of chlorhexidine. This phenomena is probably due to the fact that the more hydrophilic amino acids interact with the chlorhexidine reducing its lipophilicity. The highest effect was observed with the dicarboxylic amino acids Asp and Glu indicating that the interaction is of hydrophilic character.

We assume that the quaternary amino groups of chlorhexidine form hydrogen bonds with the carboxyl



FIGURE 4. Effect of dicarboxylic amino acids on the  $R_M$  value of chlorhexidine.

groups of the amino acids resulting in charge transfer complexes of unknown stoichiometry. Unfortunately, charge transfer chromatography does not give information about the composition of the complexes. The correlation between the concentration of dibasic amino acids in the eluent and the  $R_M$  value of chlorhexidine is markedly nonlinear (Fig.4). As in the majority of cases the correlation between the two complexing molecules is significantly linear (41,42), this finding can be tentatively explained by the assumption that the complexes may have minimally two stoichiometries TABLE 3. Effect of Dicarboxylic Amino Acids on the Lipophilicity ( $R_M$  value) of Chlorhexidine (C = Concentration of Amino Acid mM).

$$R_M = a + b.log C$$

Amino acid	n a	b	$s_{b}$	$r_{calc.}$	r <sub>99%</sub>
L-Glu	5 0.91	-0.52	0.05	0.9795	0.9587
D-Glu	5 0.99	-0.57	0.03	0.9812	0.9587
L-AsP	7 1.02	-0.73	0.02	0.9979	0.9507
D-Asp	7 1.10	-0.70	0.04	0.9813	0.9507

depending on the amino acid - chlorhexidine ratio. At low amino acid concentrations the amino acid - chlorhexidine ratio is lower in the complex than at higher concentrations resulting in the deviation from the linearity.

Calculations show that the logarithmic correlation fits well to the experimental data the significance level being in each case over 99% (Table 3).

The change of amino acid concentration accounts for about 95 - 99% of the lipophilicity change of chlorhexidine (see r values). No significant difference was found between the slope (b) values of L- and D-forms that is the interaction of chlorhexidine with Asp and Glu not stereospecific or the stereospecificity is such low that it is under the detection limit of our method. The L- and D-forms of Asp had a significantly higher effect on the  $R_M$  value of chlorhexidine than the corresponding forms of Glu (the calculated t values were 3.32 and 2.60 for L- and D-forms, respectively, the tabulated t value for 95% significance level is 2.23). It means that chlorhexidine preferably interacts with the more acidic (43) and less lipophilic (44) Asp proving again the hydrophilic character of the interaction.

Our results prove that chlorhexidine can bind to various proteins, the binding sites are probably the acidic amino acid side chains in the proteins. The interaction is of hydrophilic character and its strength depends on the type and concentration of the ions in the environment. We assume that these protein - chlorhexidine interactions may have a role in the biological activity of chlorhexidine.

This work was supported by the OTKA 2670 grant of the Hungarian Academy of Sciences.

#### REFERENCES

- Rosa, M. D. L., Sturzenberger, O. P. and Moore, D. J., The use of chlorhexidine in the management of gingivitis in children, J.Periodont.<u>59</u>, 387, 1988.
- Moran, J., Addy, M. and Newcombe, R., A clinical trial to assess the efficacy of sanguinarine-zinc mouthrinse (Veadent) compared with chlorhexidine mouthrinse (Corsodyl), J.Clin.Periodont. <u>15</u>, 612, 1988.
- Almquist, H. and Luthman, J., Gingival and mucosal reactions after intensive chlorhexidine gel treatment with or without oral hygiene measures, J.Dent.Res. <u>96</u>, 557, 1988.
- 4. Brecx, M., Liechti, T., Widmer, J., Gehr, P. and Lang, N. P., Histological and clinical parameters of human gingiva following 3 weeks of chemical (chlorhexidine) or mechanical plaque control, J.Clin.Periodont. <u>15</u>, 150, 1989.
- 5. Vignarajah, S., Newman, H. N. and Bulman, J., Pulsated jet subgingival irrigation with 0.1% chlorhexidine, simplified oral higiene and chronic periodontitis, J.Clin.Periodont. <u>15</u>, 355, 1989.

- Grossman, E., Meckel, A. H., Isaacs, R. L., Ferretti, G. A., Sturzenberger, O. P., Bollmer, B. W., Moore, D. J., Rijana, R. C. and Manhart, M. D., A clinical comparison of antibacterial mouthrinses: effects of chlorhexidine, phenolics, and sanguinarine on dental plaque and gingivitis, J.Periodont. <u>603</u>, 435, 1989.
- Giertsen, E., Scheie, A. A. and Rölla, G., Inhibition of plaque formation and plaque acidogenecity by zinc and chlorhexidine combinations, Scand.J. Dent.Res. <u>96</u>, 541, 1988.
- Schaeken, M. J. M. and de Haan, P., Effects of sustained-release chlorhexidine acetate on the human plaque dental flora, J.Dent.Res. <u>68</u>, 119, 1989.
- 9. Nuuja, T., Meurmann, J. H., Mutomaa, H., Kortelainen, S. and Metteri, J., The effect of a combination of chlorhexidine diacetate, sodium fluoride and xylitol on plaque wet weight and periodental index scores in Military Academy cadets refraining from mechanical tooth cleaning for 7-day experimental periods, J.Clin.Periodont. <u>19</u>, 73, 1992.
- 10. Meurman, J. H., Jousimies-Somer, H., Suomala, P., Alaluusua, S., Torkko, H. and Asikainen, S., Activity of amine-stannous fluoride combination and chlorhexidine against some aerobic and anaerobic oral bacteria, Oral.Microbiol.Immunol. <u>4</u>, 117, 1989.
- Kalaga, A., Addy, M. and Hunter, B., Comparison of chlorhexidine delivery by mouthwash and spray on plaque accumulation, J.Periodont. <u>50</u>, 127, 1989.
- 12. Giertsen, E., Scheie, A. A. and Rölla, G., In vivo effects of zinc and chlorhexidine on dental plaque ureolysis and glycolysis, J.Dent.Res. <u>58</u>, 1132, 1989.
- Zickert, I., Emilson, C. G., Ekblom, K. and Krasse, B., Prolonged oral reduction of Streptococcus mutans in humans after chlorhexidine disinfection followed by fluoride treatment, Scand.J. Dent.Res. <u>95</u>, 315, 1987.

- 14. Emilson, C. G., Lindquist, B. and Wennerholm, K., Recolonization of human tooth surfaces by Streptococcus mutans after suppression by chlorhexidine treatments, J.Dent.Res. <u>66</u>, 1503, 1987.
- Meurman, J. H., Ultrastructure, growth, and adherence of Streptococcus mutans after treatment with chlorhexidine and fluoride, Caries Res. <u>22</u>, 283, 1988.
- 16. Schaeken, M. J. M., de Jong, M. H., Franken, H. C. M. and van der Hoeven, J. S., Effect of chlorhexidine and iodine on the composition of the human dental plaque flora, Caries Res. <u>18</u>, 401, 1984.
- 17. Schaeken, M. J. M., van den Kiebom, C. W. A., Franken, H. C. M., de Jong, M. H. and van der Hoeven, J. S., Effects of chlorhexidine, iodine, and 5,7-dichloro-8-hydroxyquinoline on the bacterial composition of rot plaque in vivo, Caries Res. <u>18</u>, 440, 1984.
- Schaeken, M. J. M., de Jong, M. H., Franken, H. C. M. and van der Hoeven, J. S., Effects of highly concentrated stannous fluoride and chlorhexidine regimes on human dental plaque flora, J.Dent.Res. <u>65</u>, 57, 1986.
- Stanley, A., Wilson, M. and Newman, H. N., The in vitro effects of chlorhexidine on subgingival plaque bacteria, J.Clin.Periodont. <u>15</u>, 259, 1989.
- Oosterwaal, P. J. M., Mikx, F. H. M., van der Brink, M. E. and Rengli, H. H., Bactericidal concentrations of chlorhexidine digluconate, amine fluoride gel and stannous fluoride gel for subgingival bacteria tested in serum at short contact times, J.Periodont.Res. <u>24</u>, 155, 1989.
- 21. Etemadzadeh, H., Meurman, J. H., Murtomaa, H., Torkko, H., Lappi, L. and Roos, M., Effect on plaque growth and salivary microorganisms of amine flouride-stannous fluoride and chlorhexidine-containing mouthrinses, J.Clin.Periodont. <u>16</u>, 175, 1989.
- 22. Cleghorn, B. and Bowden, G. H., The effect of pH on the sensitivity of species of Lactobacillus to chlorhexidine and the antibiotics minocycline and spiramycin, J.Dent.Res. <u>68</u>, 1146, 1989.

- Netuschil, L., Reich, E. and Brecx, M., Direct measurement of the bactericidal effect of chlorhexidine on human dental plaque, J.Clin.Periodont. <u>16</u>, 484, 1989.
- 24. Ferretti, G. A., Ash, R. C., Brown, A. T., Parr, M. D., Romond, E. H. and Lillich, T. T., Control of oral mucositis and candidiasis in marrow transplantation: a prospective, double-blind trial of chlorhexidine digluconate oral rinse, Bone Marrow Transplant.<u>3</u>, 483, 1988.
- Harbison, M. A. and Hammer, S. M., Inactivation of human immunodeficiency virus by Betadine products and chlorhexidine, J.Acq.Immun.Def.Syndr. <u>2</u>, 16, 1989.
- 26. Shakespeare, V., Shakespeare, P. G. and Evans, B. T., Effect of proprietary oral rinses containing chlorhexidine, hexetidine and benzydamine on the proliferation of human buccal epithelial cells in culture, Arch.Oral.Biol. <u>33</u>, 881, 1988.
- 27. Perdok, J. F., van der Mei, H. C., Genet, M. J., Rouxhet, P. G. and Busscher, H. J., Elemental surface concentration ratios and surface free energies of human enamel after application of chlorhexidine and adsorption of salivary constituents, Caries Res. 23, 297, 1989.
- 28. Mucklow, E. S., Accidental feeding of a dilute antiseptic solution (chlorhexidine 0.05% with cetrimide 1%) to five babies, Human Toxicol. <u>7</u>, 567, 1988.
- 29. Okano, M., Nomura, M., Hata, S., Okada, N., Sato, K., Kitano, Y., Tashiro, M., Yoshimoto, Y., Hama, R. and Aoki, T., Anaphylactic symptoms due to chlorhexidine gluconate, Arch.Dermatol. <u>125</u>, 50, 1989.
- Cserháti, T., Szögyi, M. and Lelkes, L., Study of chlorhexidine-tenside interactions by means of charge-transfer chromatography, Pharm.Acta Helv. <u>65</u>, 113, 1990.
- Giertsen, E., Scheie, A. A. and Rölla, G., Antimicrobial and antiplaque effects of a chlorhexidine and Triton-X-100 combination, Scand.J.Dent. Res. <u>97</u>, 233, 1989.

- 32. Barkvoll, P., Rölla, G. and Svendsen, A. K., Interaction between chlorhexidine digluconate and sodium lauryl sulfate in vivo. J.Clin.Periodont. <u>15</u>, 593, 1989.
- 33. Chawner, J. A. and Gilbert, P., A comparative study of the bactericidal and growth inhibitory activities of the bisguanides alexidine and chlorhexidine, J.Appl.Bacteriol. <u>66</u>, 243, 1989.
- 34. Chawner, J. A. and Gilbert, P., Interaction of the bisguanides chlorhexidine and alexidine with phospholipid vesicles: evidence for separate mode of action, J.Appl.Bacteriol. <u>66</u>, 253, 1989.
- 35. Cserháti, T. and Szögyi, M., Interaction between some synthetic phospholipids and nonylphenyl-nonylglycolate studied by charge-transfer chromatography, J.Biochem.Biophys.Meth. <u>14</u>, 101, (1987).
- 36. Horváth, C., Melander, W. R. and Molnár, I., Solvophobic interactions in liquid chromatography with nonpolar stationary phases, J.Chromatogr. <u>125</u>, 129, 1976.
- 37. Cserháti, T., Lipophilicity determination of some 3,5-dinitrobenzoic acid esters on unimpregnated cellulose layer, Chromatographia <u>18</u>, 18, 1984.
- Mager, H., Moderne Regressionsanalyse, Salle, Sauerlander, Frankfurt am Main, 1982, p.135.
- 39. Klaas, E. B., Horáth, C., Melander, W. B. and Nahum, A., Surface silanols in silica-bounded hydrocarbonaceous stationary phases. II. Irregular retention behaviour and effect of silanol masking, J.Chromatogr. 203, 65, 1981.
- 40. Cserháti, T., Darwish, Y. M. and Matolcsy, G., Anomalies in the reversed-phase thin-layer chromatographic behaviour of some heterocyclic quaternary ammonium salts, J.Chromatogr. <u>241</u>, 223, 1982.
- Cserháti, T., Szögyi, M. and Györfi, L., Interaction of some nonionic tensides with dioleoylphosphatidyl choline studied by charge-transfer chromatography. J.Chromatogr. <u>349</u>, 295, 1985.

- 42. Cserháti, T. and Szögyi, M., Amino acid-amino acid interactions studied by charge-transfer chromatography, J.Chromatogr. <u>434</u>, 455, 1988.
- 43. Barrett, G. C., Chemistry and Biochemistry of the Amino Acids, Chapman and Hall, London, 1985. p.9.
- 44. Jonsson, J., Eriksson, L., Hellberg, S., Sjöström, M. and Wold, S., Multivariate parametrization of 55 coded and non-coded amino acids, Quant.Struct.-Act.Relat. <u>8</u>, 204, 1989.

Received: March 31, 1994 Accepted: January 31, 1995